

RESEARCH ARTICLE

***In-vitro* Antioxidant and Cytotoxicity (sk-mel-3 cell) Activity of Green Synthesised Copper Nanoparticle using *P. pellucida* Plant Aqueous Extract**

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ABSTRACT

Green synthesis of metallic nanoparticles using medicinal plant extract treated to environmentally friendly, stable, cost-effective. Present work to study *In-vitro* antioxidant and cytotoxicity of green synthesised CuNPs using *P. Pellucida* plant extract. Copper nanoparticles were synthesised biogenically using *P. Pellucida* plant aqueous extract. CuNPs Characterised by using FE-SEM, U.V., FT-IR, EDAX. Biogenically synthesised copper nanoparticles conducted *in-vitro* Antioxidant (DPPH) and cytotoxicity (SK-MEL-3 cell). The results CuNPs were characterised by using U.V. spectroscopy absorbance 575 nm. Scanning electronic microscope showing the distribution and shape (5-20 nm) of the nanoparticles. X-ray spectrum showed different peaks for CuNPs detected at 2θ values 35.45°, 44.32°, and 65.25°. EDAX elemental spectroscopy conformed to the copper metal inside the nanoparticle (69.7%). The synthesised CuNPs showed good free radical scavenger activity than the ascorbic acid, and it also showed significant cytotoxicity (16µg/mL) against human skin cell lines (SK-MEL-3 cell). The current study conclusion recommended that green synthesised (*P. Pellucida*) CuNPs be used for therapeutic application.

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INTRODUCTION

There are numerous techniques for the biogenic development of metallic nanomaterials. Because of their fascinating physical characteristics and possible application in various fields (medicine, pharmacy, biotechnology, chemistry etc.), technical and scientific research on the synthesis and development of metallic nanoparticles has augmented. Principally tiny size and large volumes to surface area ratio. Special attention was paid to copper nanoparticles production within metallic

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nanoparticles due to its low price of the noble metal compared to Ag, Au and Pt and their varied research and industrial (1, 2). Synthesis of nanoparticles through a green approach, non-toxic and biodegradable chemicals have been used. Nanoparticles are synthesised from different plant sources (3,4).

Medicinal plant extractions for copper nanoparticles have generous benefits, such as catalysis, photocatalytic activity (5) and bactericidal (6), DNA binding and sensors (7), anticancer (8), free radical scavenging, etc. (9, 10).

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The green synthesis of copper nanoparticles with medicinal plants extracts is biodegradable, profitable and stable (11). Biomolecules such as carbohydrates, phenols, tannins, flavonoids, and others in plant extract have been found to play a significant factor in nanocomposites reduction, aggregation and capping (12). Presently, the market for herbal supplements for health care is growing with each day.

Pellucida peperomia (shiny bush, silver bush) annual herb, and it belong to the Piperaceae family. The monsoon season grows to a height of 15-46 cm in humid soil, particularly underneath the trees (13). It is usually observed in southeast and southwest Nigeria and several tropical Asian and South American countries in the West African rainforest region. *Pellucida peperomia* used amazon region humans for the treatment of cardiac arrhythmia, diuretic, dementia disease. The leaves and stem aqueous blend is used to treat bleeding, fever, headache, stomach pain, injuries, and a cough suppressant recorded in the Ayurveda (14, 15, 16). The present study investigates in-vitro antioxidant and cytotoxicity of green synthesised copper nanoparticles using *P. pellucida* plant extract.

MATERIALS AND METHODS

Source of chemical and reagents

Cu (NO₃)₂·3H₂O, acridine orange, ethidium bromide (AO/EB), PBS (1%), DMEM (Dulbecco's Modified Eagle's Medium), streptomycin, penicillin-G, L-glutamine, phosphate-buffered saline, MTT (3-(4,5 dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide), ethylene diamine tetraacetic acid, ethanol, ethidium bromide and acridine orange (EB/AO) and DMSO (dimethyl sulfoxide), Dehydrated methanol obtained from Sigma Aldrich Chemicals Pvt. Ltd (India).

Plant collection

P. pellucida was obtained from Patambi, Kerala, India, in August 2020. A plant taxonomist (Dr Dhanapal V, Professor and Principal of Sri Sasta College of Pharmacy) authenticated the plant (Fig. 1).

Preparation of plant extract

Plant leaves are cleaned with distilled water to eliminate any particulate matter. Plant leaves make a fine powder using mortar and pestle. The extract (aqueous) was prepared using a cold maceration process. 100 g powder of *P. pellucida* Linn was saturated in 1 litre of deionised water and held in a shaker at 30 °C for 24 hrs for constant stirring at 100 rpm. It then dried out at normal temperature. The extract was then purified and processed at - 4 °C for additional study (17, 18).

Phytochemical screening

Preliminary phytochemical examination of the extract was carried out to classify active elements using basic methods (19).

Green synthesis of copper nanoparticle (CuNPs)

The aqueous plant extract was used to reduce Cu (NO₃)₂·3H₂O. The Cu (NO₃)₂·3H₂O was mixed with the aqueous plant extract and stirred continuously for at least one hr. The reaction process was repeatedly observed, and the colour change was noted. The reaction mixture was centrifuged at 12,000×g for 15 min, and the nanoparticle pellet was cleaned with distilled water (20)

Characterisation of CuNPs

CuNPs were initially studied by ultraviolet spectroscopic analysis within a 200 – 800 nm range. FTIR was used to identify the structural features and



Fig. 1. Hole plant of *P. pellucida*

selective phytochemical components. The results estimated 4000–400 cm⁻¹ range. The powder form of the nanoparticles exposed to CuKα1X-Ray diffractometer radiation ($\lambda = 1.5406 \text{ \AA}$) at 40 kV and 30 mA with 2 θ rang. CuNPs were placed on the sample holder and sputter-coated using gold, following that, the nanoparticles' average particle size and shape were studied by using TESCAN MIRA3 LMH Schottky FE-SEM (Japan) (21, 22)

Determination of antioxidant activity (CuNPs)

The ascorbic acid and CuNPs sample stock solutions prepared have a strength of 1.0 mg/ml. CuNPs and ascorbic acid concentrations of 10, 20, 40, 60, 80, 100 $\mu\text{g/ml}$, in methanol solution. CuNPs (0.5 ml), ascorbic acid transformed into 0.5ml of 0.5 mM DPPH in methanol solvent. After 30 min of storage at in the dark place at room temperature, the optical density was measured at 517 nm using Stat Fax 4200 Elisa reader (USA). Each experiment was performed in duplicate. Inhibition (I%) calculated as follows: (23)

$$\text{I\% antioxidant activity} = \frac{\text{Abs control} - \text{Abs of antioxidant}}{\text{Abs control}} \times 100$$

Cell culture maintenance

Human skin cancer cells (SK-MEL-3) were obtained from the National Centre for Cell Sciences, Pune, India. Dulbecco's Modified Eagle's medium was used to preserve the cell line, complemented by 10% of Fetal Bovine Serum. Penicillin (100 U/ml) and streptomycin (100 $\mu\text{g/ml}$) was added to prevent microbial contamination. The cell culture medium was preserved in a pressurised environment with 5 per cent CO₂ at 37°C.

Cytotoxicity activity

SK-MEL-3 cells were seeded in 96 well plates and incubated in a CO₂ incubator for 24hr to facilitate the adhesion. Cells were treated with the control and CuNPs at various concentrations and incubated cell culture incubator after 24hr cell were washed with the cell culture media and added MTT dye (Incubated 4hr 37°C). After 4hr the cells were treated with formazan and observed cell viability using a multi-well plate reader (540 nm). In comparison to the control, the proportion of stable cells was calculated. CuNPs IC₅₀ value was calculated, and the effective dose was analysed at a different period (24).

$$\text{Inhibitory of cell growth (\%)} = \frac{\text{absorbance of the control mean} - \text{absorbance of the sample Mean}}{\text{absorbance of the control Mean}} \times 100$$

IC₅₀ of nanoparticles measured from the dose-responsive curve inhibit 50% cytotoxicity compared to control cells. Experiments were carried out three times (25).

Measurement of apoptotic induction using acridine orange/ethidium bromide (AO/EB) dual staining method

Microscopic fluorescence analysis of apoptotic cell inhibition was carried out according to Liu et al. (26). SK-MEL-3 cells were seeded at 5 x 10⁴ cells/well in a 96 six-well plate and incubated for 24 hours. After treatment with CuNPs for 24 hrs, the cells were detached, washed with cold PBS and then stained with a mixture of A.O. (100 $\mu\text{g ml}^{-1}$)/E.B. (100 $\mu\text{g ml}^{-1}$) ratio (1:1) at room temperature for 5 min. Treated cells were collected and rinsed three times with PBS. The plates were stained with acridine orange/ethidium bromide (AO/EB 1:1 ratio; 100 $\mu\text{g/ml}$) for 5 minutes and examined immediately under fluorescent microscope 40x magnification. A fluorescence microscope observed the stained cells at 40x magnifications.

Statistical analysis

Results descriptive as mean \pm S.D. They restricted comparing statistical differences carried by (ANOVA) one-way analysis of variance accompanied by Duncan's Multiple Range Test, using SPSS version 12.0 for windows. Values tested for significance if the p-value was less than 0.05.

RESULTS

Phytochemical analysis

All the findings of the phytochemical investigation are shown in Table 1. Aqueous extract gave promising outcomes for steroids, terpenoids confirmed by Salkowski and Liebermann-Burchard's test in the present study. The presence of terpenoids, phenols and flavonoids were also confirmed in *P. pellucida* aqueous extract.

Evaluation of CuNPs

UV - Vis spectroscopic analysis

U.V. spectroscopy within the 200-800 nm range initially verified the development of CuNPs. A characteristic peak of 575 nm showed the absorption spectrum of green synthesised CuNPs.

Table 1. Phytochemical constituents of *P. pellucida* aqueous leaf extract.

| S. No | Phyto-Constituents | Chemical Test | Inference |
|-------|--------------------|-----------------------|-----------|
| 1 | Alkaloids | Dragendorff's reagent | + |
| 2 | Tannins | Ferric chloride | + |
| 3 | Saponins | Forthing | + |
| 4 | Flavonoids | Lead ethanoate | + |
| 5 | Steroids | Salkowski test | + |
| 6 | Terpenoids | Liebermann-Buchard's | + |
| 7 | Flavanols | Shinoda | + |
| 8 | Anthraquinones | Borntrager's | - |
| 9 | Carbohydrates | Molish's | + |

(+) present, (-) absent

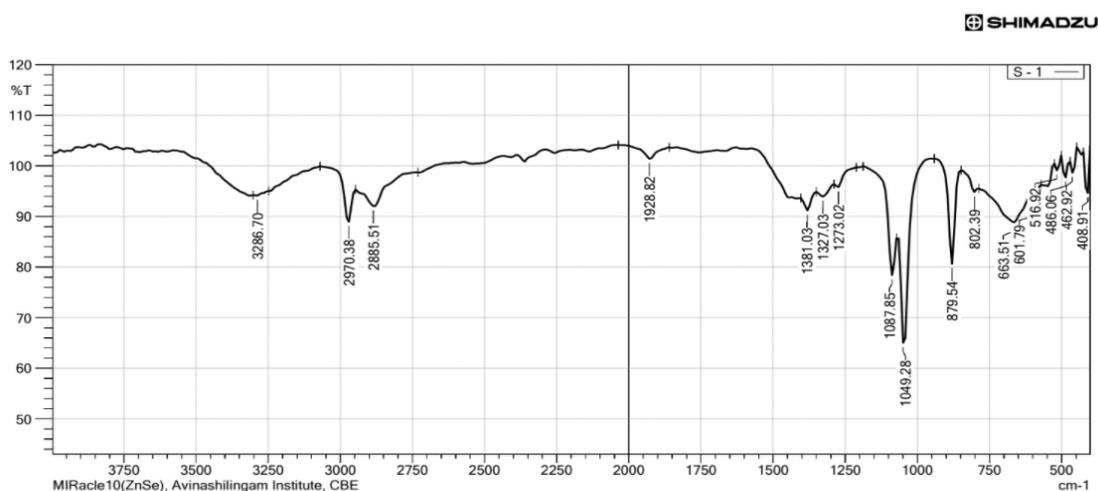


Fig. 2. FTIR analysis of CuNPs

FTIR spectroscopic analysis

A Fourier transform infrared spectroscopy was utilised as an affirmative method for the construction of nanoparticles (Fig. 2). This study offers an imprint of current molecules vibrational and rotational modes, thus helping to classify the functional and potential plant molecules involved in CuNPs nanoparticles reduction.

XRD Analysis

The structure of the nanoparticles is indicated by the XRD analysis of the synthesised CuNPs. At 2θ values of 35.45°, 44.32°, and 65.25° degrees, diffraction peaks were observed (Fig. 3).

FE-SEM analysis

The SEM study was carried out using the Schottky FE-SEM (Japan) TESCAN MIRA3 LMH model. FE-SEM analysis was used to classify the structure and scale of CuNPs. Microscopy of

CuNPs has shown that they have a nano-range particle size (500nm) spherical and homogeneous in distribution under the FE-SEM Microscope (Fig. 4).

EDAX analysis

The elemental analysis of the CuNPs from the EDAX spectrum of the FE-SEM image is shown in Fig-5. The percentage of molecular mass and atomic value of Cu respectively 69.7%. The EDAX spectrum is consistent with the presence of copper in the nanoparticle.

Antioxidant activity

The antioxidant properties of CuNPs using DPPH were evaluated and compared to ascorbic acid. The ratio of free radical inhibition of CuNPs was observed at different concentrations. The free radical inhibition visually detected colour transformation from purple to yellow suggests that

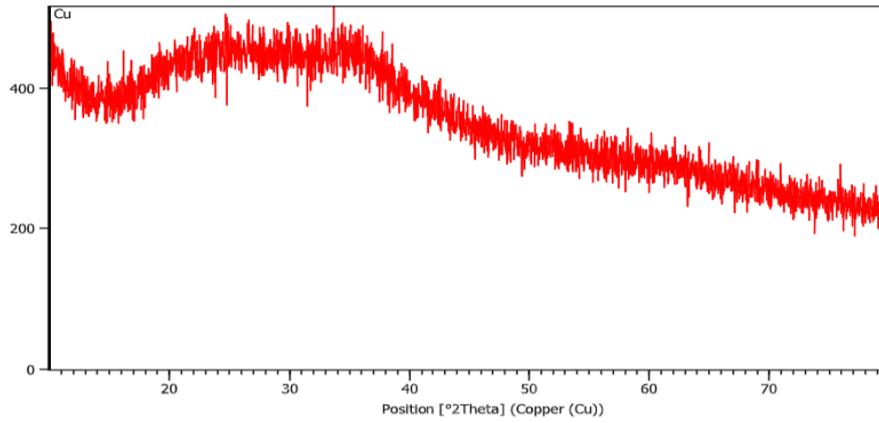


Fig. 3. XRD analysis of CuNPs

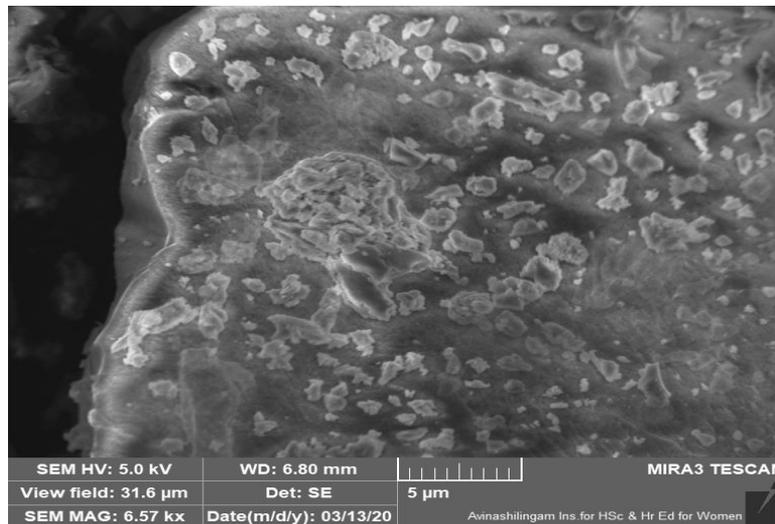


Fig. 4. FE-SEM images of CuNPs

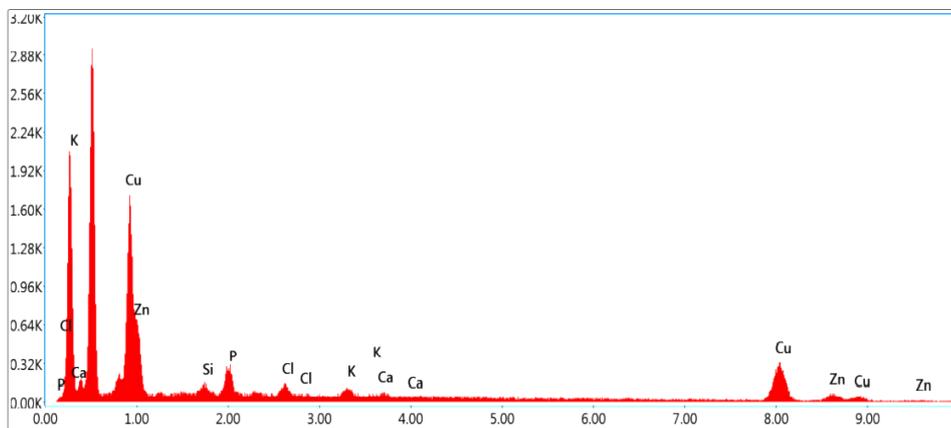


Fig. 5. EDAX analysis of green synthesised CuNPs

DPPH reduced exhibiting better scavenging action (Fig. 6).

Cytotoxic activity (MTT assay)

Cytotoxic activity against SK-MEL-3 cancer cells with different concentrations ranging from 5µg/mL, 10µg/mL, 15µg/mL, 20µg/mL, 25µg/mL and 30µg/mL tested *P. Pellucida* mediated CuNPs. The In-vitro cytotoxicity evaluation was evaluated after 48hrs. Fig 7 and 8 shows the altered structure of SK-MEL-3 cells after dose-dependent treatment with CuNPs. Compared with control cell viability, CuNPs (16µg) significantly reduced the proliferation of SK-MEL-3 cells.

DISCUSSION

In the present investigation, CuNPs were bio-synthesised from the *P. pellucida* plant extract.

Microscopical observation of CuNPs have shown that they have a particle size in the nano range; they are irregular in distribution. The dimensions, shape of the CuNPs were observed under FE-SEM. Cu particles grow slowly, form small structures.

Several investigations described biogenic preparation copper nanoparticles as environmentally approachable effective antioxidant and antitumor agents (27). The nanoparticle has a wide surface nature, permeability and biodegradability (28). Green synthesised CuNPs studied antioxidant activity using the DPPH method and showed antioxidant activity (standard ascorbic acid). Further analysis of copper nanoparticles studied cytotoxicity against SK-MEL-3 cell lines and showing good cytotoxic activity (IC50 value 16µg/mL).

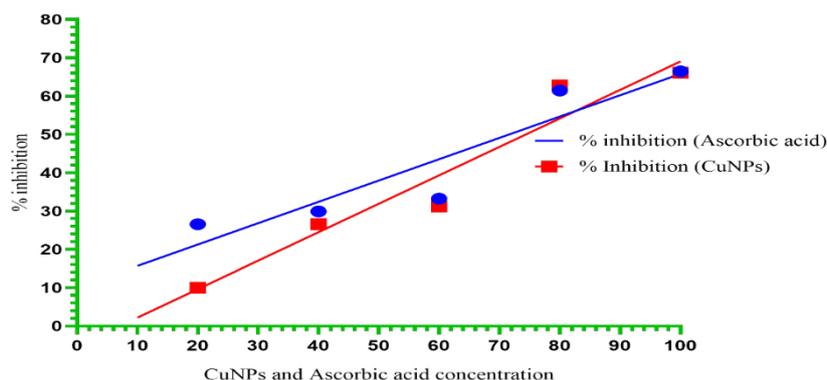


Fig. 6. Antioxidant activity of CuNPs

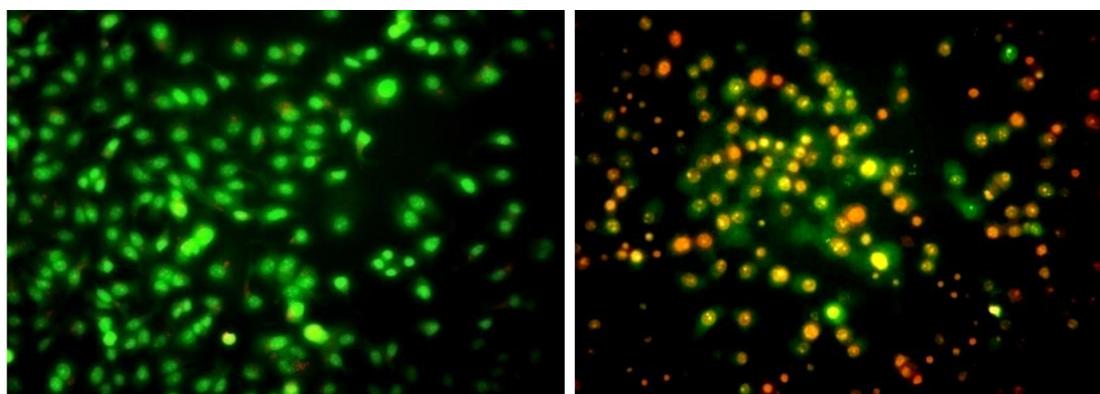


Fig. 7. Regulated human melanoma cells (SK-MEL-3) and CuNPs (16µg/mL) at 24 hrs, marked with dual stain AO/EB, were observed under fluorescence microscopy. Green nucleus appeared in living cells, an early disjointed form of the yellow nucleus with chromatin, late apoptotic condensation of chromatin of orange nuclei and necrotic cells.

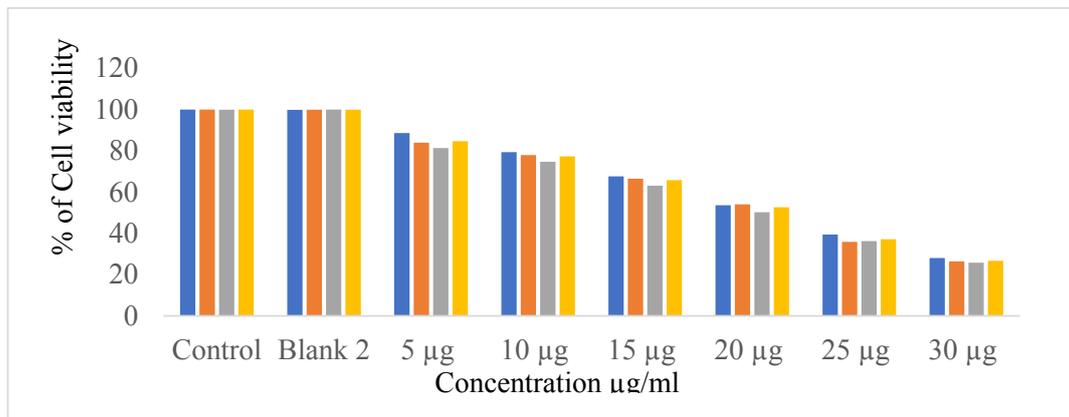


Fig. 8. Percentage of SK-MEL-3 cell viability of control and CuNPs by MTT assay at different concentrations from 5-30 µg/mL.

CONCLUSION

The convenient and environmentally friendly method to prepare CuNPs using reducing Cu (NO₃)₂·3H₂O with *P. pellucida* aqueous extract was established. *P. pellucida* aqueous extract acting as a better reducing and covering agent. Green synthesised CuNPs characterised using EDAX, U.V, FT-IR, FE-SEM and XRD techniques. The development of CuNPs shows the deviations in the colour of the solution. U.V. spectroscopy showed the 200-800 nm range initially verified the story of CuNPs. A characteristic peak of 575 nm showed the absorption spectrum of CuNPs. The crystalline nature of the synthesised nanoparticles identified by XRD analysis has diffraction values at 35.45°, 44.32°, and 65.25° degrees. FE-SEM analysis conforms to the nanoparticle size and shape. The CuNPs investigated antioxidant and cytotoxicity. The experimental results concluded that biogenic synthesised nanoparticles had significant antioxidant (DPPH) and cytotoxicity activity (SK-MEL-3 cell).

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CONFLICT OF INTEREST

Nil

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