

RESEARCH ARTICLE

Time dependent difference effects of MgO and ZnO nanoparticles on the serum and hippocampus Mg^{2+} , Zn^{2+} , $Fe^{2+/3+}$ and Ca^{2+} levels in the stressed rats

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ARTICLE INFO

Article History:

Received 7 June 2018

Accepted 26 July 2018

Published 15 August 2018

Keywords:

Elements

Hippocampus

MgO/ZnO Nanoparticles

Stress

ABSTRACT

Objective(s): Stress is a physiological response that can disrupt body elements homeostasis and lead to neurophysiological abnormality. This study has been investigated the serum and hippocampus Mg^{2+} , Zn^{2+} , $Fe^{2+/3+}$ and Ca^{2+} level changes in two times after MgO NPs and ZnO NPs single injection following restraint stress in the male rat.

Methods: Animals were divided into two main groups that each of them includes: control, restraint of 90, 180 and 360 min+ saline, MgO NPs and ZnO NPs 5 mg/kg alone and with a restraint of 90 min. In one group, 30 min and in another 120 min after intraperitoneally injections of components or stress induction elements levels were measured in the serum/ hippocampus.

Results: Elements level changed in the serum and hippocampus following injections of MgO and ZnO NPs depend on acute time after injections. ZnO NPs induced a positive correlation between serums $Fe^{2+/3+}$ levels in two acute times. Different times of stress induction have different effects on elements level changes in the serum and hippocampus, 30 and 120 min after induction and nanoparticles could alleviate these changes depend on the time. In restraint groups, there were positive and negative significant correlations between two different times measurements of $Fe^{2+/3+}$ or Ca^{2+} in the serum and hippocampus.

Conclusion: it seems that time is an important factor in ameliorative MgO NPs and ZnO NPs effects on elements disruption induced by stress, but their exact interaction with stress systems containing ions level changes needs to more investigation.

How to cite this article

Torabi M, Kesmati M, Pourreza N, Najafzadeh Varzi H, Galehdari H. Time dependent difference effects of MgO and ZnO nanoparticles on the serum and hippocampus Mg^{2+} , Zn^{2+} , $Fe^{2+/3+}$ and Ca^{2+} levels in the stressed rats. *Nanomed Res J*, 2018; 3(4): 197-205. DOI: 10.22034/nmrj.2018.04.004

INTRODUCTION

Acute and chronic immobilization stress can affect elements homeostasis and change their distribution in the different parts of the body, especially in the brain [1, 2]. Immobilization stress was decreased endogenous Zn^{2+} and $Fe^{2+/3+}$

concentrations in different parts of the brain such as hippocampus 24 hours after the stress induction [3]. On the other hand, trace elements such as Mg^{2+} , Zn^{2+} , $Fe^{2+/3+}$ and Ca^{2+} are essential for living cells and body systems functions[4-6] and can change balance of each other in the body[7,8]. Magnesium

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and zinc have key roles in the central nervous system and hippocampus functions [9, 10].

Both of them work on some similar receptors, like N-methyl-D-aspartate (NMDA), that block or reduce its activity [11]. It has been shown that in rats under Zn^{2+} restriction Mg^{2+} , $Fe^{2+/3+}$ and Ca^{2+} levels in the serum have increased significantly and decreased Zn^{2+} concentration in the serum and hippocampus, while could not affect Mg^{2+} , $Fe^{2+/3+}$ and Ca^{2+} levels in the rat's hippocampus [12]. Magnesium oxide and zinc oxide nanoparticles (MgO NPs and ZnO NPs), as novel sources of Mg^{2+} and Zn^{2+} , are widely used in medicine and pharmacology, by the development of nanotechnology [13, 14]. Benefit and toxic effects of these components have been investigated in the various living cells and systems, because of the unique properties of them [13, 15-18].

Some studies have shown that nanoparticles can change ions level in the central nervous system and neural cells [16, 19, 20]. Amara et al. (2013) have shown that in the rat brain ZnO NPs change elements levels including $Fe^{2+/3+}$, Zn^{2+} , and Ca^{2+} [19]. Also, ZnO NPs increase intracellular Ca^{2+} level and affect neural cell by increasing Zn^{2+} ions [16, 20]. Ion releasing is one of the most important cytotoxic factors of metal oxide nanoparticles and their primary particle size or surface area did not affect cellular functions directly [13].

Ben-Slama and et al. (2015) have indicated that oral exposure to ZnO NPs decreased the brain Ca^{2+} concentration [21]. Our previous study has shown that ZnO NPs can increase Zn^{2+} level in the serum and decrease anxiety-like behaviors in animal models [22].

In this work we investigated and compared

the MgO NPs and ZnO NPs effect on Mg^{2+} , Zn^{2+} , $Fe^{2+/3+}$, and Ca^{2+} level changes in the serum/hippocampus of adult male rats at two different acute times following acute stress induction.

MATERIAL AND METHODS

Animals grouping and treatments

In this experimental work, male Wistar rats (220 \pm 10 g) were purchased from animal house of faculty of veterinary in the Shahid Chamran University of Ahvaz. Experiments were carried out under ethical code of EE/96.24.3.88369/scu.ac.ir. MgO and ZnO nanoparticles (USnano., CO, USA) (Fig. 1), suspensions were prepared before experiments and injected intraperitoneally in a single dose of 5 mg/kg and a volume of 1 mL/kg [22, 23]. Both nanoparticles didn't form large aggregates that blocked the syringe during injections. Rats were divided into 16 groups, which included two sub-groups, 1) non-restraint rats: control, MgO NPs and ZnO NPs 5 mg/kg and 2) restraint rats: restrained for 90, 180 and 360 min + saline (ST 90 min, ST 180 min and ST 360 min) and ST 90 min + MgO or ZnO NPs 5 mg/kg. In all restraint groups, components were injected immediately after restraint stress induction, then in one main group (including, 8 groups) 30 min and in the other (including, 8 groups) 120 min after components injections or restraint stress induction animals were killed for a measure of ions concentrations in the serum and hippocampus. The number of rats in each group was six.

Acute stress induction

Rats were restrained for 90, 180 and 360 min in the plexiglass tubes, then immediately received saline (1 ml/kg) or MgO NPs and ZnO NPs 5 mg/kg.

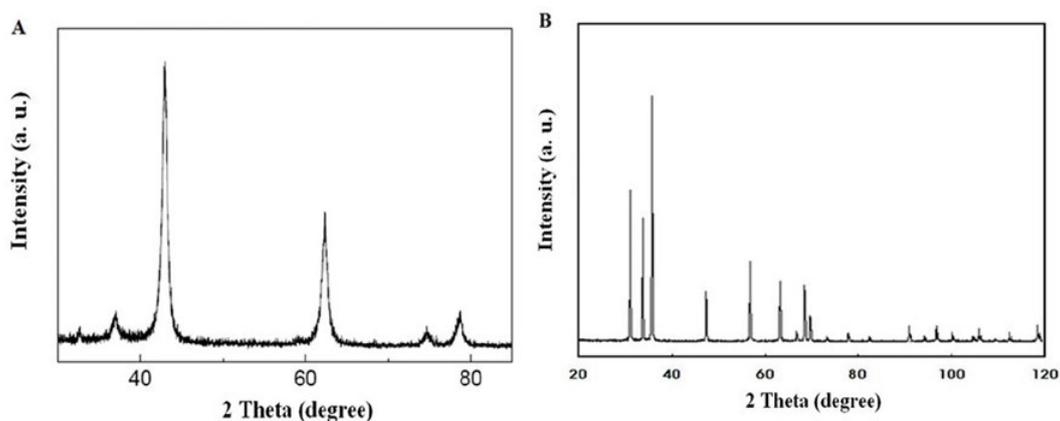


Fig. 1. XRD patterns of MgO NPs (A) and ZnO NPs (B).

Serum/ hippocampus sampling and assessment of elements contents

In the first 8 groups after 30 min and in the second 8 groups after 120 min, all rats scarified then serums and hippocampus homogenates of them obtained. Elements contents measured by a flame atomic absorption spectrophotometer apparatus in all samples, and results expressed as a $\mu\text{g}/\text{mL}$ of the serum and mg/g of wet hippocampus tissue.

Statistical analysis

One way ANOVA with Tukey post-hoc was used for comparing among groups and Student's t-test was used for comparing the means of unpaired data by using SPSS 16 software. Pearson correlation coefficient was calculated between elements contents of the serum/ hippocampus in 2 acute different times. Differences with a p value of <0.05 was considered statistically significant. Results are presented as the mean \pm standard error of the mean and graphs are plotted with the Excel software.

RESULTS AND DISCUSSION

Nanoparticles size detection by XRD patterns

Fig. 1 is XRD patterns of the MgO NPs (A) and ZnO NPs (B) and indicates that the sizes of both nanoparticles were lower than 100 nm before injections.

Assessment of Mg^{2+} concentration

As seen in Fig. 2A, MgO NPs (30 min ($P=0.002$) and 120 min ($P<0.0001$)) and ZnO NPs (30 min ($P<0.001$) and 120 min ($P=0.003$)) significantly increased Mg^{2+} level in the serum 30 and 120 min after injections. Also, in MgO NPs group Mg^{2+} level 120 min after injection was significantly higher than 30 min ($P=0.0018$). Level of Mg^{2+} in serum was increased 30 min after ST 90 ($P=0.0064$) and ST 180 min induction ($P=0.038$) and was decreased and reached to the control group at 120 min. ST 360 min did not affect the level of Mg^{2+} significantly after 30 and 120 min compared to the control groups but was seen significant difference between

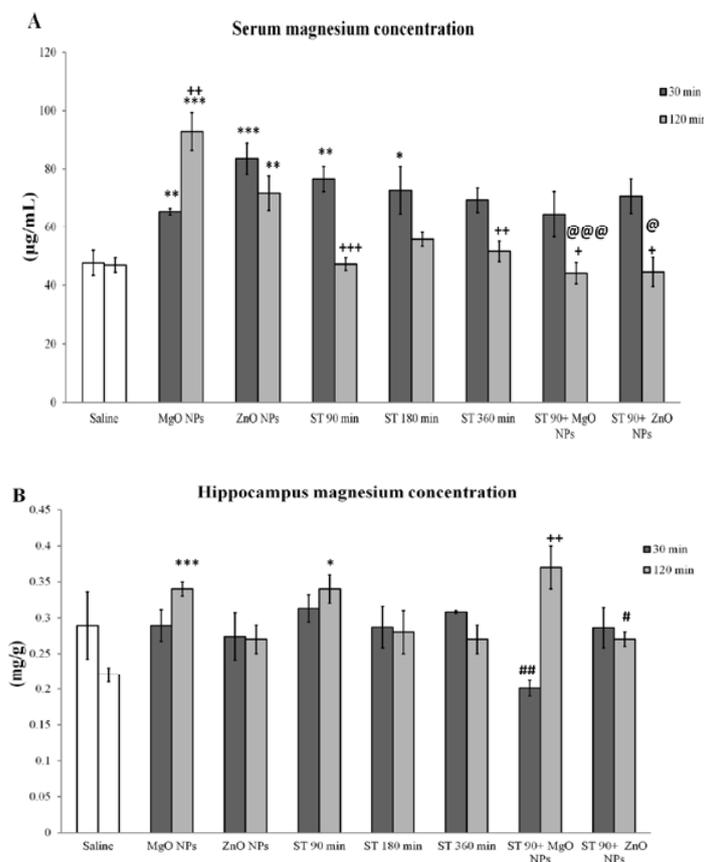


Fig. 2. Assessment of Mg^{2+} concentration. * $P<0.05$, ** $P<0.01$ and *** $P<0.001$ are in comparison with the control group at the same time (30 and 120 min). + $P<0.05$, ++ $P<0.01$ and +++ $P<0.001$ are between equal treatment groups in two different times. # $P<0.05$ and ## $P<0.01$ are in comparison with ST 90 min groups at the same time (30 and 120 min). @ $P<0.05$ and @@@ $P<0.001$ are in comparison with MgO NPs or ZnO NPs groups at the same time (30 and 120 min). All bars are means \pm standard error of the means.



30 and 120 min (P=0.0092).

In the restraint groups, MgO NPs and ZnO NPs did not change Mg^{2+} level in comparison with ST 90 min group after 30 min and 120 min, but following injection of both nanoparticles Mg^{2+} level after 120 min was decreased in comparison with after 30 min (P=0.042). Data analysis showed that ST 90 min has a reductive effect on the MgO NPs (P<0.0001) and ZnO NPs (P=0.0167) effects on serum magnesium concentration 120 min after injection.

Fig. 3B showed that MgO NPs (P=0.0004) and ST 90 min (P=0.041) increased hippocampus magnesium level after 120 min. While MgO NPs acted significantly in opposite directions in the presence of ST 90 min at two different times of 30 and 120 min (P=0.0030). MgO NPs in the presence of stress after 30 min reduced magnesium compared with the administration of MgO NPs alone but increased it after 120 min. This effect was

not observed for ZnO NPs.

These results show that time duration after MgO NPs injection is an important factor in the release of Mg^{2+} from it in the serum and hippocampus. Also, stress has a dual role in the release of Mg^{2+} , in a short time can increase Mg^{2+} in the serum and while with passing the time increase it in the hippocampus.

Assessment of Zn^{2+} concentration

MgO NPs significantly decreased Zn^{2+} level after 120 min (P=0.0002), while ZnO NPs increased Zn^{2+} level in the serum after 30 (P=0.013) and 120 min (P=0.040). Stress was decreased Zn^{2+} level in a time-dependent manner 120 min after induction (ST 90 min (P<0.05), ST180 min (P<0.01) and ST 360 min (P<0.001)). In the restraint groups, MgO NPs (P=0.0075) and ZnO NPs (P=0.030) have increased Zn^{2+} level after 30 min, while decreased it after 120

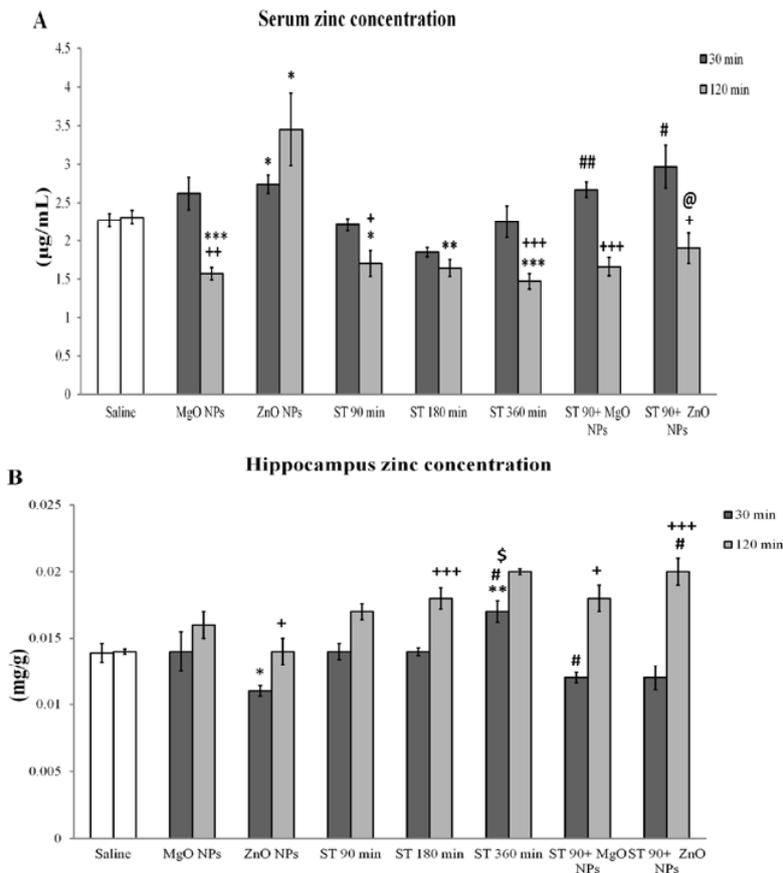


Fig. 3. Assessment of Zn^{2+} concentration. *P<0.05, **P<0.01 and ***P<0.001 are in comparison with the control group at the same time (30 and 120 min). +P<0.05, ++P<0.01 and +++P<0.001 are differences between equal treatment groups in two different times (30 and 120 min). #P<0.05 and ##P<0.01 are in comparison with ST 90 min groups at the same time (30 and 120 min). \$P<0.05 shows a significant difference in comparison with ST 180 min group at the same time (30 min). @P<0.05 shows difference in comparison with ZnO NPs group at the same time. All bars are means ± standard error of the means.

min in comparison with 30 min after injections (MgO NPs ($P<0.001$) and ZnO NPs ($P<0.05$)). Stress 90 min decreased the serum zinc level in the ZnO NPs group ($P=0.013$) after 120 min that show negative effect of stress on ZnO NPs efficacy especially (Fig. 3A).

ZnO NPs was decreased Zn^{2+} level in the hippocampus after 30 min ($P=0.015$). Stress increased partially Zn^{2+} level 30 and 120 min after induction in all groups and it was significant 30 min after induction of ST 360 min ($P=0.0058$). In restraint groups, MgO NPs and ZnO NPs have increased Zn^{2+} level after 120 min and it was significant in the ZnO NPs group ($P=0.044$) (Fig. 3B).

These results show that except in one group (ZnO NPs) in all the other groups decrease of Zn^{2+} in serum was parallel to increase of it in the hippocampus, 120 min after treatments.

Assessment of $Fe^{2+/3+}$ concentration

MgO NPs increased serum $Fe^{2+/3+}$ level in the stressed ($P<0.0001$) and non-stressed ($P=0.0005$) animals after 30 min, while ZnO NPs decreased $Fe^{2+/3+}$ level partially. Iron level decreased after 120 min in ST 360 min group ($P=0.011$). In the restraint animals MgO NPs significantly decreased $Fe^{2+/3+}$ level after 120 min in comparison with 30 min ($P<0.0001$) (Fig. 4A).

MgO NPs ($P<0.0076$) and ZnO NPs ($P=0.0018$) significantly increased $Fe^{2+/3+}$ level in the hippocampus of non-restraint animals after 30 min. Also stress increased $Fe^{2+/3+}$ level in all duration after 30 min (ST 90 and 180 min ($P<0.001$) and ST 360 min ($P<0.05$)) and decreased it after 120 min. In the restraint animals MgO NPs decreased $Fe^{2+/3+}$ level after 120 min in compared with 30 min ($P=0.0012$). On the other hand, ZnO

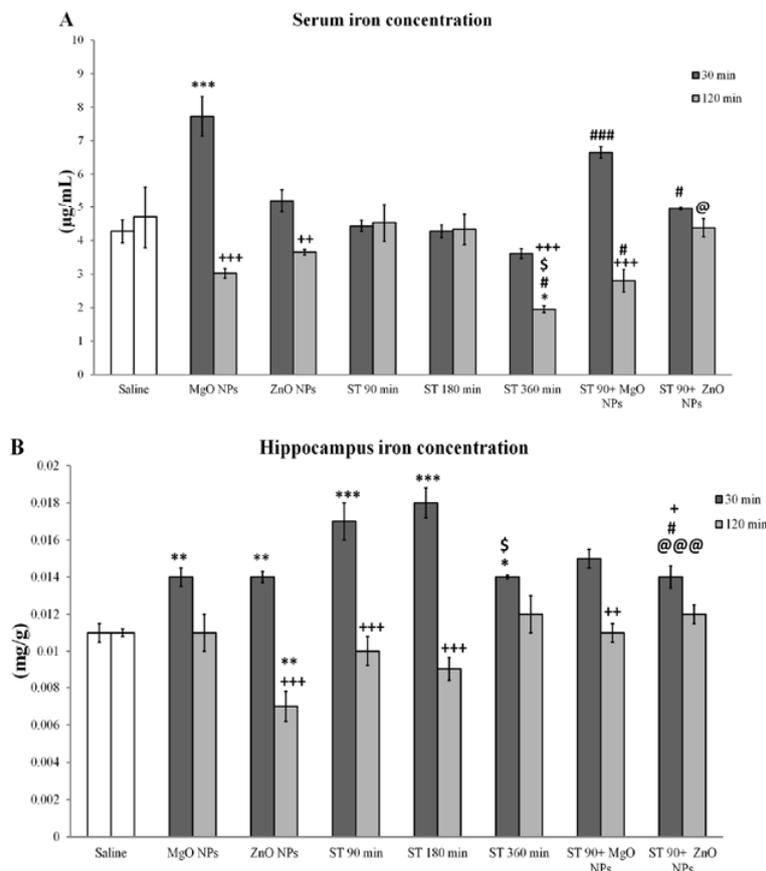


Fig. 4. Assessment of $Fe^{2+/3+}$ concentration. * $P<0.05$, ** $P<0.01$ and *** $P<0.001$ are in comparison with the control group at the same time (30 and 120 min). + $P<0.05$, ++ $P<0.01$ and +++ $P<0.001$ are differences between equal treatment groups in two different times (30 and 120 min). # $P<0.05$ and ### $P<0.001$ are differences in comparison with ST 90 min groups at the same time (30 and 120 min). \$ $P<0.05$ is a significant difference in comparison with ST 180 min group at the same time (30 and 120 min). @ $P<0.05$ and @@@ $P<0.001$ are differences in comparison with ZnO NPs group at the same time. All bars are means \pm standard error of the means.

NPs significantly decreased $Fe^{2+/3+}$ level in the non-restraint ($P=0.0031$) after 120 min (Fig. 4B).

According to these results, it seems that acute injection of both nanoparticles could change the $Fe^{2+/3+}$ balance in two of measurement sites and stress influences their effects.

Assessment of Ca^{2+} concentration

In the serum, MgO NPs increased Ca^{2+} concentration after 30 min ($P=0.0106$), while ZnO NPs increased it after 120 min ($P=0.012$). The ST 90 min significantly decreased Ca^{2+} in 120 and 30 min after induction ($P<0.01$), while ST 360 min increased it and was significant after 30 min ($P<0.05$). In the restraint animals, MgO NPs was increased Ca^{2+} level after 30 min and decreased it after 120 min ($P<0.0001$), while in these animals ZnO NPs was increased Ca^{2+} level in both times after injection (30 min ($P<0.0001$) and 120 min ($P=0.0021$)). Also, in MgO NPs ($P=0.0002$) and

ZnO NPs recipients groups stress had negative effect on calcium level and decreased it in comparison with nanoparticles injections alone after 120 min ($P=0.0106$).

MgO NPs significantly was increased Ca^{2+} level in the hippocampus, after 120 min ($P=0.008$), while ZnO NPs increased it after 30 min ($P=0.0054$) (Fig. 5 A and B). Stress in all duration increased Ca^{2+} level after 30 min and it was significant in the ST 180 min group ($P=0.037$). In the restraint animals, MgO NPs decreased Ca^{2+} level after 30 min ($P=0.0105$), while ZnO NPs decreased it after 120 min ($P=0.0042$) (Fig. 5B).

Based on these findings, MgO NPs and ZnO NPs effects on Ca^{2+} level balance, in the restraint and non-restraint rats, were completely adverse in two different acute times.

Assessment of Pearson correlation between ions concentration in two acute times (30 and 120 min) Data on Table 1 show that there were significant

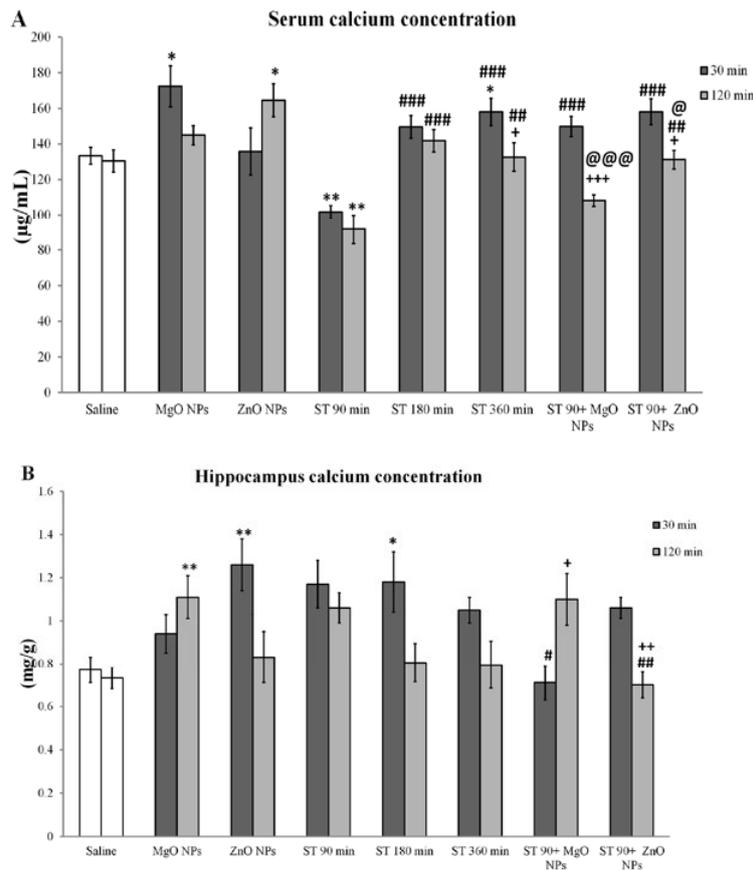


Fig. 5. Assessment of Ca^{2+} concentration. * $P<0.05$ and ** $P<0.01$ are differences in comparison with the control group at the same time (30 and 120 min). + $P<0.05$, ++ $P<0.01$ and +++ $P<0.001$ are differences between equal treatment groups in two different times. # $P<0.05$, ## $P<0.01$ and ### $P<0.001$ are differences in comparison with ST 90 min groups at the same time (30 and 120 min). @ $P<0.05$ and @@@ $P<0.001$ are differences in comparison with MgO NPs or ZnO NPs groups at the same time. All bars are means \pm standard error of the means.

Table 1. Pearson correlation between elements contents at the two times

A												
In the serum												
Groups	Saline		ST 90, 180 and 360 min+ saline		MgO NPs		ZnO NPs		ST 90 min+ MgO NPs		ST 90 min+ ZnO NPs	
	R	P	R	P	R	P	R	P	R	P	R	P
Elements												
Mg ²⁺	0.672	0.14	-0.14	0.574	0.258	0.621	0.221	0.674	0.316	0.541	0.211	0.689
Zn ²⁺	0.754	0.08	-0.29	0.242	0.424	0.402	-0.700	0.122	0.087	0.869	0.065	0.903
Fe ^{2+/3+}	-0.39	0.44	0.607	0.008**	0.441	0.381	-0.673	0.143	-0.015	0.978	-0.700	0.122
Ca ²⁺	-0.52	0.28	0.626	0.005**	-0.532	0.277	0.225	0.669	-0.569	0.238	-0.509	0.303

B												
In the hippocampus												
Groups	Saline		ST 90, 180 and 360 min+ saline		MgO NPs		ZnO NPs		ST 90 min+ MgO NPs		ST 90 min+ ZnO NPs	
	R	P	R	P	R	P	R	P	R	P	R	P
Elements												
Mg ²⁺	0.417	0.41	0.186	0.461	0.39	0.442	0.230	0.661	0.392	0.442	0.356	0.488
Zn ²⁺	0.147	0.78	0.043	0.865	0.30	0.556	0.214	0.684	-0.060	0.910	0.170	0.748
Fe ^{2+/3+}	-0.57	0.22	-0.47	0.04*	-0.79	0.057	-0.313	0.545	-0.370	0.471	0.661	0.153
Ca ²⁺	0.275	0.59	-0.16	0.511	0.80	0.052	0.844	0.035*	-0.550	0.258	0.471	0.346

*= $P < 0.5$ and **= $P < 0.01$

positive correlations between both of $Fe^{2+/3+}$ ($R=0.0607$, $P=0.008$) and Ca^{2+} ($R=0.626$, $P=0.005$) level changes in the serum, 30 and 120 min after stress induction. There was a negative correlation between $Fe^{2+/3+}$ content of the hippocampus in two different times after stress induction ($R=-0.471$, $P=0.049$). Also, in ZnO NPs group there was a positive correlation between Ca^{2+} level changes in the hippocampus, at two different acute times ($R=0.844$, $P < 0.035$).

Results have indicated that efficacy of MgO and ZnO NPs on ions level changes in the serum and hippocampus could be different with passing the time. Since metal oxide nanoparticles are dissolved easily in acidic environments, probably ZnO NPs can dissolve in the lysosomes and release Zn^{2+} ions [18, 24, 25]. This is a possible way to increase of Mg^{2+} and Zn^{2+} in the serum during the times after nanoparticles injections. Oxide salts are less reactive and release of ions from them is slow [26]. Previously we have indicated that Zn^{2+} concentration increased in the serum of male rats following injection of ZnO NPs and conventional ZnO and retention of Zn^{2+} ions in the serum of ZnO NPs group was higher than conventional ones after 24 hours; so that probably in the rat body ZnO NPs clearance was less than conventional forms [22]. This retention of nanoparticles in the body can affect their efficacy with passing the time.

Injections of nanoparticles could change the balance of elements too. Magnesium is a natural Ca^{2+} antagonist and can regulate Ca^{2+} channels with an important role in the active transport of Ca^{2+} ions through the cell membranes [27, 28]. On the

other hands, there is a divalent metal transporter 1 that transports divalent metals including Mg^{2+} , Zn^{2+} and Ca^{2+} by a proton-coupled mechanism [29]. Some Zn^{2+} transporter proteins can facilitate non-transferrin bound $Fe^{2+/3+}$ -mediated delivery in cultured cells and similar trans membrane pores conduct $Fe^{2+/3+}$ and Ca^{2+} through the membranes [6]. Consumption of a Ca^{2+} supplement decreased the total $Fe^{2+/3+}$ absorption, primarily by reducing the initial uptake of heme $Fe^{2+/3+}$ [30]. Transferrin receptor (TfR)-mediated $Fe^{2+/3+}$ transport by the blood-brain-barrier and $Fe^{2+/3+}$ concentration is high in the hippocampus of the normal brain and TfR in the cerebral endothelial of the hippocampus is about 3–7 folds higher than in the cortex [31].

All of these studies indicated that MgO NPs and ZnO NPs could affect the balance of other elements in the serum and hippocampus, it's while maybe nanoparticles directly change the balance of elements, that this needs too more investigation. At the following results have indicated that restraint stress has different effects on elements changes in the serum and hippocampus depend on the acute time passing after stress induction.

Karakoc and et al. (2003) have shown that acute immobilization stress causes endogenous Zn^{2+} release from the brain and may enhance production of the brain iron transport proteins [1]. In all over the world iron deficiency anemia is a popular nutritional deficiency anemia and it has been reported that Zn^{2+} supplementation prevents stress effects and a stress-induced decrease in $Fe^{2+/3+}$ level [32, 33]. However, usage of a modest Zn^{2+} supplement induce a cellular $Fe^{2+/3+}$ deficiency and

probably further reduce of the $Fe^{2+/3+}$ statue [34].

Also, it has been shown that in the rat hippocampus stress increases Ca^{2+} current amplitude [35]. In this study MgO NPs and ZnO NPs could improve $Fe^{2+/3+}$ and Ca^{2+} concentration changes in the serum and hippocampus following stress induction and their effects depend on the acute time passing after injection.

CONCLUSION

It seems that rather than the elements level changes by nanoparticles, the efficacy of MgO and ZnO NPs on ions level imbalance induced by restraint stress depend on the acute time passing after stress induction. But more investigation needs to find exact effects of nanoparticles on body ions balance in healthy and stressful situations as well as in different acute and chronic times.

ACKNOWLEDGMENT

This study is supported financially by the Research Council of the Shahid Chamran University of Ahvaz (Grant: 96/3/02/16670).

CONFLICT OF INTEREST

The authors declare that there are no conflicts of interest.

REFERENCES

- Karakoc Y, Yurdakoc E, Gulyasar T, Mengi M, Barutcu UB. Experimental stress-induced changes in trace element levels of various tissues in rats. *The Journal of Trace Elements in Experimental Medicine*. 2003;16(1):55-60.
- Teng W-f, Sun W-m, Shi L-f, Hou D-d, Liu H. Effects of Restraint Stress on Iron, Zinc, Calcium, and Magnesium Whole Blood Levels in Mice. *Biological Trace Element Research*. 2007;121(3):243-8.
- Kamal Z, Najimi M, Chigr M, El Ouahli M, Er-Raoui G, Chigr F. Trace elements distribution in the brain of stressed rats. *American Journal of Neuroscience*. 2012; 3(2): 79-86.
- Feng H, Guo L, Gao H, Li X-A. Deficiency of calcium and magnesium induces apoptosis via scavenger receptor BI. *Life Sciences*. 2011;88(13-14):606-12.
- Li W, Yu J, Liu Y, Huang X, Abumaria N, Zhu Y, et al. Elevation of brain magnesium prevents synaptic loss and reverses cognitive deficits in Alzheimer's disease mouse model. *Molecular Brain*. 2014;7(1).
- Pantopoulos K, Porwal SK, Tartakoff A, Devireddy L. Mechanisms of Mammalian Iron Homeostasis. *Biochemistry*. 2012;51(29):5705-24.
- Baltaci AK, Mogulkoc R, Belviranli M. Serum levels of calcium, selenium, magnesium, phosphorus, chromium, copper and iron--their relation to zinc in rats with induced hypothyroidism. *Acta Clinica Croatica*. 2013; 52(2):151-156.
- Bicer M, Akil M, Sivrikaya A, Kara E, Baltaci AK, Mogulkoc R. Effect of zinc supplementation on the distribution of various elements in the serum of diabetic rats subjected to an acute swimming exercise. *Journal of Physiology and Biochemistry*. 2011;67(4):511-7.
- Yang Y, Jing X-P, Zhang S-P, Gu R-X, Tang F-X, Wang X-L, et al. High Dose Zinc Supplementation Induces Hippocampal Zinc Deficiency and Memory Impairment with Inhibition of BDNF Signaling. *PLoS ONE*. 2013;8(1):e55384.
- Yorulmaz H, Şeker FB, Demir G, Yalçın İE, Öztaş B. The Effects of Zinc Treatment on the Blood-Brain Barrier Permeability and Brain Element Levels During Convulsions. *Biological Trace Element Research*. 2012;151(2):256-62.
- Sowa-Kućma M, Szewczyk B, Sadlik K, Piekoszewski W, Trela F, Opoka W, et al. Zinc, magnesium and NMDA receptor alterations in the hippocampus of suicide victims. *Journal of Affective Disorders*. 2013;151(3):924-31.
- Doboszewska U, Szewczyk B, Sowa-Kućma M, Noworyta-Sokołowska K, Misztak P, Gołębiowska J, et al. Alterations of Bio-elements, Oxidative, and Inflammatory Status in the Zinc Deficiency Model in Rats. *Neurotoxicity Research*. 2015;29(1):143-54.
- Horie M, Fujita K, Kato H, Endoh S, Nishio K, Komaba LK, et al. Association of the physical and chemical properties and the cytotoxicity of metal oxide nanoparticles: metal ion release, adsorption ability and specific surface area. *Metallomics*. 2012;4(4):350.
- Teymuri Zamaneh H, Kesmati M, Malekshahi Nia H, Najafzadeh Varzi H, Torabi M. Investigating the effects of chronic magnesium oxide nanoparticles on aerobic exercise-induced antinociception in adult male rats. *International Journal of Green Pharmacy*. 2017; 11(4) (Suppl) S892.
- Ghobadian M, Nabiani M, Parivar K, Fathi M, Pazooki J. Toxic effects of magnesium oxide nanoparticles on early developmental and larval stages of zebrafish (*Danio rerio*). *Ecotoxicology and Environmental Safety*. 2015;122:260-7.
- Karmakar A, Zhang Q, Zhang Y. Neurotoxicity of nanoscale materials. *Journal of Food and Drug Analysis*. 2014;22(1):147-60.
- Moeini-Nodeh S, Rahimifard M, Baeri M, Abdollahi M. Functional Improvement in Rats' Pancreatic Islets Using Magnesium Oxide Nanoparticles Through Antiapoptotic and Antioxidant Pathways. *Biological Trace Element Research*. 2016;175(1):146-55.
- Zhang J, Qin X, Wang B, Xu G, Qin Z, Wang J, et al. Zinc oxide nanoparticles harness autophagy to induce cell death in lung epithelial cells. *Cell Death and Disease*. 2017;8(7):e2954.
- Amara S, Slama IB, Omri K, Ghoul JEL, Mir LEL, Rhouma KB, et al. Effects of nanoparticle zinc oxide on emotional behavior and trace elements homeostasis in rat brain. *Toxicology and Industrial Health*. 2013;31(12):1202-9.
- Huang Y-F, Liu H, Xiong X, Chen Y, Tan W. Nanoparticle-Mediated IgE-Receptor Aggregation and Signaling in RBL Mast Cells. *Journal of the American Chemical Society*. 2009;131(47):17328-34.
- Ben-Slama I, Mrad I, Rihane N, EL Mir L, Sakly M, Amara S. Sub-acute oral toxicity of Zinc Oxide nanoparticles in male rats. *Journal of Nanomedicine and Nanotechnology*. 2015; 6(3): 1-6.
- Torabi M, Kesmati M, Harooni HE, Varzi HN. Different Efficacy of Nanoparticle and Conventional ZnO in an Animal Model of Anxiety. *Neurophysiology*. 2013;45(4):299-305.

23. Kesmati M, Zadehdarvish F, Jelodar Z, Torabi M. Vitamin C potentiate sedative effect of magnesium oxide nanoparticles on anxiety and nociception in the postpartum depression model. *Nanomedicine Journal*. 2017; 4(1): 17-24.
24. Bannunah AM, Vllasaliu D, Lord J, Stolnik S. Mechanisms of Nanoparticle Internalization and Transport Across an Intestinal Epithelial Cell Model: Effect of Size and Surface Charge. *Molecular Pharmaceutics*. 2014;11(12):4363-73.
25. Cho W-S, Duffin R, Howie SEM, Scotton CJ, Wallace WAH, MacNee W, et al. Progressive severe lung injury by zinc oxide nanoparticles; the role of Zn²⁺ dissolution inside lysosomes. *Particle and Fibre Toxicology*. 2011;8(1):27.
26. Veldkamp T, van Diepen JTM, Bikker P. The bioavailability of four Zn²⁺ Oxide Sources and Zn²⁺ sulfate in broiler chickens; Lelystad, Wageningen UR (University & Research center) LivestockResearch, Confidential Livestock Research Report. 2014; 806 pages: 27
27. Gröber U, Schmidt J, Kisters K. Magnesium in Prevention and Therapy. *Nutrients*. 2015;7(9):8199-226.
28. Romani AMP. Cellular magnesium homeostasis. *Archives of Biochemistry and Biophysics*. 2011;512(1):1-23.
29. Nadadur SS, Srirama K, Mudipalli A. Fe^{2+/3+} transport & homeostasis mechanisms: Their role in health & disease. *Indian Journal of Medical Research*. 2008; 128: 533-544.
30. Roughead ZK, Zito CA, Hunt JR. Inhibitory effects of dietary calcium on the initial uptake and subsequent retention of heme and nonheme iron in humans: comparisons using an intestinal lavage method. *The American Journal of Clinical Nutrition*. 2005;82(3):589-97.
31. Zheng W, Monnot AD. Regulation of brain iron and copper homeostasis by brain barrier systems: Implication in neurodegenerative diseases. *Pharmacology & Therapeutics*. 2012;133(2):177-88.
32. Li Y, Zheng Y, Qian J, Chen X, Shen Z, Tao L, et al. Preventive Effects of Zinc Against Psychological Stress-Induced Iron Dyshomeostasis, Erythropoiesis Inhibition, and Oxidative Stress Status in Rats. *Biological Trace Element Research*. 2012;147(1-3):285-91.
33. Saboor M, Qamar K, Qudsia F, Khosa SM, Moinuddin M, Usman M. Malabsorption of iron as a cause of iron deficiency anemia in postmenopausal women. *Pakistan Journal of Medical Sciences*. 2015;31(2).
34. Donangelo CM, Woodhouse LR, King SM, Viteri FE, King JC. Supplemental Zinc Lowers Measures of Iron Status in Young Women with Low Iron Reserves. *The Journal of Nutrition*. 2002;132(7):1860-4.
35. Joels M, Velzing E, Nair S, Verkuyl JM, Karst H. Acute stress increases calcium current amplitude in rat hippocampus: temporal changes in physiology and gene expression. *European Journal of Neuroscience*. 2003;18(5):1315-24.